

0957-4166(95)00450-5

Diastereoselectivity in the Bakers Yeast Reduction of [1-2H](Sorbaldehyde)Fe(CO)3

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Abstract: Bakers yeast reduction of the faster reacting (2S) enantiomer of [1-2H](sorbaldehyde) Fe(CO)3 proceeds with complete diastereoselectivity. Little or no diastereoselectivity is observed in bakers yeast reduction of the slower reacting (2R) enantiomer or in the reduction of the racemic complex with NaBH4.

INTRODUCTION

Biotransformations using bakers yeast have received much attention in recent years, particularly (but not exclusively) in the asymmetric reduction of carbonyl compounds.¹ Though several exceptions have been reported, the reactions may generally be viewed as proceeding according to Prelog's rule² via hydrogen transfer to the *re* face of a prochiral carbonyl with R_L and R_S representing large and small substituents adjacent to the carbonyl group.



For ketones containing chiral substituents, bakers yeast reductions have been widely used for resolution in both organic¹ and, to a lesser extent, in organometallic systems.³ Such resolutions depend either on differences in the rate of reduction of the enantiomers of the racemate, or since the *re* selectivity is essentially maintained for both enantiomers, formation of separable diastereoisomers from which the optically pure ketones can be regenerated by oxidation. In the case of non-deuteriated aldehydes, such diastereoisomer formation is not possible, and relatively few examples of kinetic resolution of organic aldehydes have been reported;⁴ the use of bakers yeast for resolution of organometallic aldehydes having planar chirality is more extensive.⁵ Though several achiral organic⁶ and organometallic^{3b}, ^{5b}, ⁷ deuteroaldehydes have been reduced with high enantioselectivity to (S)-RCHD(OH) derivatives, we are not aware of any study of the

relative diastereoselectivity of bakers yeast reduction of a chiral organometallic or organic deuterioaldehyde. Following our successful kinetic resolution of (sorbaldehyde)Fe(CO)3 using bakers yeast, 5^{c} we wish to report here our results on the diastereoselectivity of the reduction of (2S)- and (2R)-[1-2H](sorbaldehyde)Fe(CO)3 which demonstrate the importance of both planar chirality and aldehyde configuration in the diastereoselection process.

RESULTS AND DISCUSSION

The $[1-^{2}H]$ derivative 4 was prepared by reduction of methyl sorbate 1 with LiAlD4 (98 %) to give 2 followed by complexation and Swern oxidation to give racemic 4. To avoid any ambiguity in the bakers yeast reduction, 4a,b was resolved at this stage by treatment with either (-)- or (+)-ephedrine.⁸ Fractional crystallisation provided the enantiomers 5a,b of defined absolute configuration⁹ from which the enantiomerically pure deuterioaldehyde was regenerated by mild hydrolysis.

Consistent with our results on kinetic resolution, 5^{e} the (2S) enantiomer is rapidly consumed (4 hours) by bakers yeast to yield the alcohol **6a** whose NMR spectrum in the H1 region contains only a single doublet resonance, consistent with complete diastereoelectricity in the reduction process. In contrast, consumption of **4b** is much slower (50 % after 15 hours) and the NMR spectrum in the H1 (Figure 1) region contains two doublets of equal intensity assignable to **6b** and **7b**, indicating no diastereoselection in the reduction process.

Figure 1. ¹H NMR Spectra of Monodeuteriated Alcohols and Acetates (C6D6)



(a) 6b/7b from BY reduction of 4b(c) 8a from BY reduction of 4a

(b) 8b/9b from BY reduction of 4b(d) 8a,b/9a,b from NaBH4 reduction of 4a,b



Scheme 1

Spectra which are sharper (due to the absence of O-H coupling) and with a greater chemical shift difference may be obtained by conversion to the acetates 8a and 8b/9b (Figure 1), though integration and coupling

constants clearly show a reversal of the chemical shift ordering of Ha and Hb. Molecular modelling studies,¹⁰ coupled with several crystallographic studies,¹¹ indicate that the OH or OAc group will occupy the least sterically hindered position shown below, perpendicular to the diene plane. Ha and Hb will thus differ in chemical shift and coupling to H2.



The (S)-C1 stereochemistry assigned to **6a** is consistent with reduction of other deuteriated aldehydes by bakers yeast, and has been confirmed by *in situ* protonation of the acetate **8a** using HSO₃F/CD₂Cl₂ at low temperature. Literature data¹² on protonation of **7** and **8** with methyl in place of deuterium show that protonation is stereospecific at low temperature, and extension to **8a** predicts that protonation/elimination should yield the initial intermediate *trans*-pentadienyl cation **10a** which rapidly undergoes bond rotation to give the *cis*-cation **11a**. The high field part of the ¹H NMR spectrum of the resultant cation **11a** is illustrated in Figure 2a, together in 2b with that of a racemic mixture of **11a,b** and **12a,b** produced by protonation of a 1:1 mixture of the racemic acetates **8a,b/9a,b** derived *inter alia* from NaBH4 reduction of **4a,b** (*vide infra*). In Figure 2b, two doublet resonances are seen at 2.05 and 3.51 ppm assignable to the inner (Hi) and outer (Ho) terminal dienyl protons respectively¹² of the **11a,b/12a,b** mixture. In Figure 2a, the low field doublet is entirely absent, indicating the presence of only **11a**. This cation is configurationally stable, exhibiting no deuterium scrambling between Hi and Ho over a period of hours at +10 °C.



NMR studies clearly show that [1-1H] analogues of 4 exist in solution in an s-*cis/s-trans* equilibrium with respect to rotation about C1-C2.9,13



Though the barrier for s-cis/s-trans interconversion is too low to measure by NMR for the tricarbonyl complex, low temperature spectra for Fe(CO)₂PR₃ complexes⁹ establish J₁₋₂ values of ≤ 2 Hz for the s-cis isomer and 8 Hz for the s-trans isomer. For the [1-¹H] analogue of 4, the average value of J₁₋₂ in MeOD at 25 °C is 4.7 Hz, indicating an approximately equal concentration of both conformers. If Prelog's rule is obeyed, the results for 4a thus indicate preferential reaction with only the s-trans conformer in which the re face of the aldehyde is exposed to attack. For 4b, in which the re face is exposed in the s-cis conformer, no diastereoselectivity is observed. Thus, lack of recognition of planar chirality is accompanied by a lack of discrimination between conformational isomers. A similar diastereoselectivity is seen in the asymmetric allylboration of (sorbaldehyde)Fe(CO)3¹⁴ where the faster and slower reacting enantiomers also exhibit greatly different diastereoselectivities in the creation of the new chiral centre.

These results may be compared to the NaBH4 reduction of racemic 4a,b where the spectrum of the derived acetate (Figure 1) indicates only a slight excess of 6a,b relative to 7a,b. The spectrum of the acetate obtained by NaBD4 reduction of (1-1H)(sorbaldehyde)Fe(CO)3 is identical but with the intensities of the Ha/Hb resonances reversed. Diastereoselectivity in this case broadly reflects the position of the conformational equilibrium.

Finally, it may be noted that coordination to iron protects the dienal from further double bond reduction by bakers yeast which occurs in the free ligand series.¹⁵ In addition, literature results¹⁶ indicate that nucleophilic displacement of acetate from 8a should occur with complete retention of configuration, thus providing access to a wider range of enantiomerically pure CHD derivatives.

EXPERIMENTAL

NMR spectra were recorded using a JEOL GSX270 spectrometer; chemical shifts are relative to tetramethylsilane. Methyl sorbate¹⁷ and $[1-^{1}H](sorbaldehyde)Fe(CO)3^{8b}$ were prepared by literature methods. Preparative thin layer chromatography was performed on a Harrison Research 7924 Chromatotron using 2 mm silica gel (type PF60254) plates. Diethyl ether was distilled from LiAlH4. Pyridine was distilled from KOH. DMSO and Et3N were distilled from CaH2 and stored over molecular seives under argon. CH2Cl2 was distilled from P2O5 under argon. Oxalyl chloride was distilled under argon and stored over molecular seives. Bakers yeast was obtained locally in Paris.

(a) **Preparation of [1-2H](sorbaldehyde)Fe(CO)3 4**

A suspension of LiAlD4 (98 %) (1.0 g, 23.8 mmol) in diethylether (30 ml) was cooled to -78 °C under nitrogen. With stirring, a solution of methyl sorbate (2.67 g, 23.8 mmol) in diethyl ether (30 ml) was added dropwise and the reaction was allowed to warm to room temperature. On completion (as judged by tlc), water (30 ml) was added followed by 10 % H₂SO₄ to dissolve the precipitate. After separation, the aqueous layer was extracted with chloroform (3 x 40 ml) and the combined organic extracts were washed with 10 % NaHCO3 solution and dried over MgSO₄. Removal of solvent gave [1-²H₂]2,4-hexadien-1-ol **2** as an oily solid (2.0 g, 85 %). NMR (CDCl₃): 5.6 - 6.3 (m, H₂-5), 1.72 (dd, Me, J₅₋₆ = 6.6, J₄₋₆ = 1.2).

Compound 2 (1.0 g, 10.2 mmol) and Fe₂(CO)9 (10.9 g, 30 mml) were stirred in toluene (30 ml) under argon in an ultrasonic bath until completion of reaction as judged by tlc. Diethyl ether (20 ml) was added and the suspension was filtered through celite. After removal of solvent, the crude product was purified by column chromatography (alumina IV, 4:1 petroleum ether (30-40): ethyl acetate) to give 3 as a brown oil (1.37 g, 56 %). A spectroscopic sample was purified by short-path distillation (125 °C, 0.1 mm Hg). Infrared (hexane): 2043, 1981, 1971 cm⁻¹; NMR (CDCl₃): 5.10 (m, H3,4), 1.04 (d, H2, J₂₋₃ = 8.4), 1.21 (m, H5, J₅₋₆ = 6.0), 1.38 (d, H6); the CH₂ resonance at 3.60 of the $[1^{-1}H_2]$ complex is absent.

DMSO (0.39 ml, 5.06 mmol) in CH₂Cl₂ (1.0 ml) was added under argon to a stirred solution of oxalyl chloride (0.23 ml, 2.53 mol) in CH₂Cl₂ (5 ml) at -78 °C. After 10 minutes stirring, complex 3 (0.569 g, 2.3 mmol) in CH₂Cl₂ (0.5 ml) was added dropwise and the reaction was stirred at -78 °C for 25 minutes. NEt₃ (1.6 ml) was added and the reaction was allowed to warm to 10 °C. After addition of water (15 ml) and further CH₂Cl₂ (15 ml), the mixture was shaken vigorously to produce an orange/yellow two phase solution. Separation and extraction of the aqueous phase (CH₂Cl₂, 3 x 100 ml), followed by drying over MgSO4 and

removal of solvent gave a brown oil which was purified by chromatotron (4:1 petroleum ether (30 - 40): ethyl acetate) to give 4 as a golden oil (0.25 g, 45 %). Infrared (hexane): 2059, 1999, 1983 cm⁻¹; NMR (C6D6): 0.66 (H2, d, $J_{2-3} = 8.1$), 5.08 (m, H3), 4.26 (m, H4), 0.83 (m, H5, $J_{5-6} = 6.5$), 1.48 (d, H6); the CHO resonance at 9.0 of the [1-¹H] complex is absent.

(b) Resolution of 4a,b

Details are given for the $[1^{-1}H]$ analogue of 4. (Sorbaldehyde)Fe(CO)3 (0.88 g, 3.73 mmol) and (-)ephedrine (0.62 g, 3.73 mmol) were stirred in dichloromethane at room temperature under N₂ until infrared monitoring indicated complete consumption of starting material (ca. 2 hours). The mixture was filtered through celite and the solvent removed. Fractional crystallisation from petroleum ether (60-80) gave yellow crystals of the $[1^{-1}H]$ analogue of 5a (0.49 g, 34 %) as a yellow solid. M.p. 123-124 °C; analysis: calc. C-59.6, H-5.52, N-3.66 %, found C-59.8, H-5.60, N-3.49 %; infrared (hexane): 2047, 1983, 1975 cm⁻¹; NMR (CDCl₃): 3.35 (d, H1, J₁₋₂ = 8.5), 0.97 (t, H2, J₂₋₃ = 7.5), 5.18 (m, H3, J₃₋₄ = 5.4), 5.02 (m, H4, J₄₋₅ = 8.0), 1.26 (m, H5, J₅₋₆ = 5.8), 1.37 (d, H6), 4.94 (d, H7, J₇₋₈ = 8.1), 2.79 (m, H8, J₈₋₁₀ = 6.6), 2.23 (s, H9), 0.57 (d, H10). Diastereoisomeric purity was assessed using either H1 or H7, which in the crude diastereoisomer mixture exhibit well distinguished additional doublet resonances at 3.72 and 4.88 ppm respectively. Treatment of **4a,b** with (-)- and (+)-ephedrine in a similar way provides **5a** and **5b** whose NMR spectra are identical and differ only from the $[1^{-1}H]$ analogue in the absence of H1 resonances and the absence of H1-H2 coupling (doublet rather than triplet resonance for H2).

(c) Hydrolysis of 5a,b

Details are given for the $[1^{-1}H]$ analogue of 5a. Water (0.3 g) and 70-235 mesh silica gel (3 g) were mixed and suspended in CH₂Cl₂ (7 ml). Complex 5a (1.0 g, 2.61 mmol) was added and the reaction monitored to completion by infrared (ca. 4 hours). Filtration and evaporation of solvent gave (+)-(2S)-(sorbaldehyde)Fe(CO)₃ (0.55 g, 89 %) which was shown to be enantiomerically pure (within the limits of ¹H detection) by examination of the CHO resonance in C₆D₆ in the presence of tris [heptafluoropropylhydroxymethylene)-(+)-camphorato] europium (III); (+)-(2S): $[\propto]_D^{20}$ +110 (c=1, CHCl₃), (-)-(2R): $[\propto]_D^{20}$ -110 (c=1, CHCl₃). Enantiomerically pure samples of the [1-²H] analogues 4a and 4b were obtained in the same way through hydrolysis of 5a and 5b respectively.

Hydrolysis of the residue from fractional recrystallisation provides material of about 70 % enrichment in the opposite enantiomer.

(d) Bakers yeast reduction of 4a/4b

Water (100 ml, degassed with argon), bakers yeast (10 g) and glucose (2.5 g) were stirred at 25 °C for 30 minutes. Complex 4a (0.28 g, 1.18 mmol) in distilled ethanol (2 ml) was added with stirring. The monitoring indicated complete consumption after 4 hours. The product was extracted with diethylether (3 x 500 ml), dried with MgSO4, and the solvent removed to give 6a as a yellow oil (0.23 g, 81 %). Infrared (hexane): 2052, 1980, 1972 cm⁻¹; NMR (C6D6): 4.31 (m, H4), 4.50 (m, H3), 0.70 (m, H2, H5), 1.01 (d, H6, J5-6 = 6.1), 3.15 (d, H1b, J1-2 = 7.4).

Reduction of 4b in the same manner using bakers yeast proceeded only to 50 % completion over 15 hours. The NMR spectrum is identical to that of 6a, but with the addition of a doublet resonance due to H1a

at 3.28 ppm ($J_{1-2} = 5.2$). The 6b/7b mixture was converted to the acetate 8b/9b [see part (f)] and then separated from unreacted 4b by chromatotron.

(e) NaBH4 reduction of 4a,b

NaBH4 (0.016 g, 0.422 mmol) was added with stirring at 0 °C under nitrogen to a solution of racemic 4 (0.1 g, 0.42 mmol) in dry methanol (10 ml). After 30 minutes, water (15 ml) and diethyl ether (20 ml) were added. After separation, the aqueous layer was extracted with diethyl ether (2 x 30 ml) and the combined extracts dried over MgSO4. Removal of solvent gave 6a,b/7a,b (80 mg, 79 %).

NaBD4 (98 %) reduction of [1-¹H(sorbaldehyde)Fe(CO)3 was carried out in the same way. NMR spectra are identical to those of 6a,b/7a,b, differing only in the relative intensities of the H1a/H1b resonances.

(f) Esterification of 6a

Complex 6a (0.23 g, 0.96 mmol) was dissolved in dry pyridine (3.5 ml) under nitrogen and cooled to 0 °C. Acetic anhydride (0.102 ml, 1.08 mmol) was added and the reaction stirred at room temperature overnight. After removal of solvent, the residue was purified by chromatotron (4:1 petroleum ether (30-40)/ethyl acetate) to give 8a as a yellow oil (0.19 g, 70 %). Infrared (hexane): 2043, 1983, 1975 cm⁻¹; NMR (C₆D₆): 4.58 (m, H3), 4.28 (m, H4), 0.6-0.8 (m, H2,H5), 0.96 (d, H6, J₅₋₆ = 5.5), 1.69 (s, OAc), 3.99 (d, H1b, J₁₋₂ = 7.8); $[\propto]_D^{20}$ -17 (c=1, CHCl3). Examination of the acetate singlet in the presence of tris[heptafluoropropylhydroxy-methylene)-(+)-camphorato]europium (III) showed no loss of optical purity of planar chirality during the bakers yeast reduction.

Esterification of **6b/7b** in the same way gave **8b/9b** having an identical NMR spectrum, but with the addition of a doublet resonance due to H1a at 3.79 ppm ($J_{1-2} = 5.8$). Esterification of the products of NaBH4/NaBD4 reduction of [1-2H]/(1-1H)(sorbaldehyde)Fe(CO)3 gave acetates having identical NMR spectra, differing only in the relative intensities of the H1a/H1b resonances.

(g) In situ Protonation

The acetate (10-15 mg) was dissolved in CD₂Cl₂ (1.5 ml) in a 5 mm NMR tube, degassed with nitrogen and cooled to -80 °C. After addition of HSO₃F (0.1 ml, triply distilled) by syringe, the solution was mixed by shaking and placed in the NMR spectrometer and warmed to -20 °C.

References

- 1. Csuk, R.; Glanzer, B. Chem. Rev., 1991, 91, 49-97.
- 2. Prelog, V. Pure Appl. Chem., 1964, 9, 119-130.
- (a) Gillois, J.; Buisson, D.; Azerad, R.; Jaouen, G. J. Organomet. Chem., 1989, 367, 85-93; (b)
 Yamazaki, Y.; Uebayasi, M.; Hosono, K. Eur. J. Biochem., 1989, 184, 671-680; (c) Yamazaki, Y.;
 Kobayashi, H. Tetrahedron: Asymmery, 1993, 4, 1287-1294.
- see, for example, (a) Ehrler, J.; Giovannini, F.; Lamatch, B.; Scebach, D. Chimia, 1986, 40, 172-173;
 (b) Zuger, M.F.; Giovannini, F.; Seebach, D. Angew. Chem., Int. Ed. Engl., 1983, 22, 1012; (c) Kawahara, K.; Mutsumoto, M.; Hashimoto, H.; Miyano, S. Chem. Lett., 1988, 1163-1164; (d) Izumi, T.; Hinata, T. J. Chem. Technol. Biotechnol. 1992, 55, 227-231.
- (a) Yamazaki, Y.; Uebayasi, M.; Someya, J.; Hosono, K. Agric. Biol. Chem., 1990, 54, 1781-1789;
 (b) Izumi, T.; Murakami, S.; Kasahara, A. Chem. Ind., 1990, 79-80;
 (c) Yamazaki, Y.; Hosono, K.;

Matsuda, H.; Minomi, N.; Asai, M.; Nakanishi, H. Biotech. Bioen., **1991**, 38, 1218-1222; (d) Top, S.; Jaouen, G.; Baldoli, C.; del Buttero, P.; Maiorana, S. J. Organomet. Chem., **1991**, 413, 125-135; (e) Howell, J.A.S.; Palin, M.G.; Jaouen, G.; Top, S.; el Hafa, H., Cense, J.M. Tetrahedron: Asymmetry, **1993**, 4, 1241-1252; (f) Izumi, T.; Hino, T.; Ishihara, A. J. Chem. Technol. Biotechnol., **1993**, 56, 45-49; (g) Izumi, T.; Hino, T.; Kasahara, A.; J. Chem. Technol. Biotechnol., **1991**, 50, 571-573.

- (a) Crombie, L.; Heavers, A.D. J. Chem. Soc., Perkin Trans. 1, 1992, 1929-37; (b) Mosher, H.S. Tetrahedron, 1974, 30, 1733-1745; (c) Liggero, S.H.; Sustmann, R.; Schleyer, P. von R. J. Am. Chem. Soc., 1969, 91, 4571-4573; (d) Althouse, V.E.; Feigl, D.M.; Sanderson, W.A.; Mosher, H.S. J. Am. Chem. Soc., 1966, 88, 3595-3599; (e) Loewus, F.A.; Westheimer, F.H.; Vennesland, B. J. Am. Chem. Soc., 1953, 75, 5018-5023; (f) Nagai, U.; Kobayashi, J.I. Tetrahedron Lett., 1976, 2873-2874; (g) Guenther, H.; Alizade, M.A.; Kellner, M.; Biller, F.; Simon, H. Z. Naturforsch., 1973, 28C, 241-246.
- 7. Sokolov, V.I.; Troitskaya, L.L.; Reutov, O.A. Dokl. Akad. Nauk SSR, 1977, 237, 1376-1379.
- (a) Djedani, F.; Grée, R.; Martelli, J.; Grée, D.; Leroy, L.; Bolard, J.; Toupet, L.; *Tetrahedron Lett.*. 1989, 30, 3781-3784; (b) Howell, J.A.S.; Bell, A.G.; O'Leary, P.J.; McArdle, P.; Cunningham, D.; Stephenson, G.; Hastings, M. Organometallics, 1994, 13, 1806-1812.
- 9. Howell, J.A.S.; Squibb, A.D.; Bell, A.G.; McArdle, P.; Cunningham, D.; Goldschmidt, Z.; Gottlieb, H.E.; Hezroni-Langerman, D.; Grée, R. Organometallics, 1994, 13, 4336-4351.
- 10. Clinton, N.A.; Lillya, C.P. J. Am. Chem. Soc., 1970, 92, 3058-3064.
- (a) Riley, P.E.; Davis, R.E.; Acta Crystallogr., 1976, 32B, 381-386; (b) Gree, D.M.; Martelli, J.T.; Gree, R.L.; Toupet, L.J. J. Org. Chem., 1995, 60, 2316-2317; (c) Lellouche, J.P.; Breton, P.; Beaucourt, J.P.; Toupet, L.J.; Gree, R. Tetrahedron Lett., 1988, 29, 2449-2452.
- (a) Lillya, C.P.; Sahatjian, R.A. J. Organomet. Chem., 1970, 25, C67-C70; (b) Sorensen, T.S.; Jablonski, C.R. J. Organomet. Chem., 1970, 25, C62-C66.
- 13. Howell, J.A.S.; Walton, G.; Tirvengadum, M.C.; Squibb, A.D.; McArdle, P.; Palin, M.G.; Cunningham, D.; Goldschmidt, Z.; Gottlieb, H.E. J. Organomet. Chem., 1991, 401, 91-123.
- 14. Roush, W.R.; Park, J.C. Tetrahedron Lett., 1990, 33, 4707-4710.
- 15. Gramatica, P.; Manitto, P.; Monti, D.; Speranza, G. Tetrahedron, 1988, 44, 1299-1304.
- (a) Uemura, M.; Minami, T.; Yamashita, Y.; Hiyoshi, K.; Hayashi, Y. Tetrahedron Lett., 1987, 28, 641-644; (b) Roush, W.R.; Wada, C.K. Tetrahedron Lett., 1994, 35, 7347-7350.
- 17. Elvidge, J.A.; Linstead R.P.; Orkin, B.A.; Sims, P.; Baer, H.; Pattison, D.B. J. Chem. Soc., 1950, 2228-2235.

(Received in UK 28 November 1995)